

CLINICAL PRACTICE

Point-of-care coagulation testing and transfusion algorithms

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Patients with cardiovascular disease have an array of haemostasis disorders that predispose to the development of thrombotic and embolic disease states. These patients are often maintained on anti-thrombotic medication to prevent adverse cardiovascular events. Patients undergoing cardiac surgery also have haemostatic disorders that include their intrinsic disease state, adjunctive medication, and the coagulation disturbances induced by cardiopulmonary bypass. The following review introduces the monitors that are available for monitoring perioperative coagulation, with an emphasis on cardiovascular surgery. Heparin monitors, platelet function monitors for use in transfusion algorithms, and monitoring anti-platelet drugs will be discussed.

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Patients undergoing coronary artery bypass grafting (CABG) account for \sim 10% of the estimated 3.2 million annual recipients of red blood cell transfusions. 41 The haemostatic management of patients undergoing cardiac surgery with cardiopulmonary bypass (CPB) necessitates an important and delicate balance between anticoagulation during CPB and proper haemostasis after CPB. These patients are at risk for excessive perioperative blood loss often requiring transfusion of blood products. In the past, clinicians administered blood products empirically due to long turnaround times of laboratory-based coagulation tests. This practice exposes patients to inappropriate blood therapy resulting in increased morbidity and mortality and hospital costs.³ 10 65 Today, point-of-care (POC) devices are available providing rapid bedside monitoring to aid the clinician in directing appropriate targeted therapy. ⁵⁹ 62 The majority of POC devices used today can perform multiple coagulation tests. The use of transfusion algorithms in conjunction with POC testing has been shown to reduce both transfusion requirements and blood loss in cardiac surgery. ^{1 9 15} 17 25 27 29 56 66 69 This review discusses current and pertinent perioperative POC monitors for coagulation, platelet function, and anti-platelet drug therapy and their respective roles in transfusion-guided algorithms.

Platelet dysfunction in cardiac surgery

Patients who present for cardiac surgery often have pre-existing platelet defects that can be acquired or drug-induced. Many patients are prescribed anti-thrombotic medication for disease states such as peripheral vascular disease and cerebrovascular disease, and thus can present for surgery with pharmacologically impaired coagulation and/or platelet function.²⁸ Platelet number can also be reduced in patients who have been exposed to heparin. The potential for relative thrombocytopenia and platelet dysfunction makes platelet function very vulnerable. CPB itself also has many anti-platelet adverse effects. 36 The literature supports that CPB down-regulates glycoprotein (GP)Ib and GPIIbIIIa receptors and decreases platelet responsiveness to thrombin and adenosine diphosphate (ADP). ²³ ⁴⁴ ⁵⁵ ⁷⁰ ⁷⁶ After cardiac surgery utilizing CPB, platelet function is compromised for at least 24 h. Couple this dysfunction with pre-existing anti-platelet effects from medication, and it becomes evident why it is so important to monitor platelet function in cardiac surgery. 26 67 POC platelet analysers provide rapid assessment of platelet function and can measure the effects of anti-platelet therapy.

Platelet function is a complex series of interactions of the endothelium with whole blood that provides platelets and coagulation factors for haemostasis. The gold standard measure of platelet function is platelet aggregometry using platelet-rich plasma. This laboratory technique is labour-intensive and time-consuming and thus is not applicable for the surgical patient. POC tests of platelet function have become more prevalent in monitoring both surgical and medical patients. Because these tests assay whole blood, are portable, and are user-friendly, they are more easily

adaptable for use at the bedside. All platelet function tests are not alike; the aetiology of the platelet dysfunction dictates that the type of test can be used to monitor platelet function. Platelet inhibition in patients treated with anti-thrombotic drugs can be tested using a variety of tests as discussed below. After CPB, the platelet defect is quite profound and is best monitored with visco-elastic whole-blood tests such as thromboelastography without modification.

Platelet function can be assessed in the static phase, dynamic phase, and as a response to an agonist. Static measures of platelet function, such as the measure of β-thromboglobulin release or mean platelet volume, are not very valuable measures of platelet function, as they capture function at a single point in time. Dynamic tests, such as the visco-elastic tests and the response to a platelet agonist, are more reflective of platelet function over time. The specific agonist used can be thrombin or collagen which test the platelet response to endothelial injury and thus its ability to mediate haemostasis in vivo. These agonists are commonly used for platelet function tests as utilized in transfusion algorithms for bleeding patients. Other agonists can be used to assess platelet responsiveness in patients taking anti-thrombotic drugs. The commonly used tests use ADP or arachidonic acid in order to test the efficacy of clopidogrel or aspirin, respectively, in inhibiting platelet function.

POC platelet function tests using a platelet agonist

Platelet Function Analyzer (PFA-100[®])

The Platelet Function Analyzer (PFA-100) (Siemens, Deerfield, IL, USA) monitor conducts a modified quantitative in vitro bleeding time under artificially created highshear conditions. Whole blood is placed on a test cartridge and a vacuum perfuses the blood across a collagen-coated membrane. An aperture is created in the membrane by a 'punch' in the presence of either epinephrine or ADP as agonist. The shear force of whole blood being drawn through a vacuum activates platelets and promotes platelet adherence and aggregation. The time it takes for a clot to form inside the glass tube and prevent further blood flow is measured as closure time.³⁸ Measurements of closure time depend on functional platelet GPIb and GPIIbIIIa, von Willebrand factor, platelet count, and haematocrit.³⁹ The response to epinephrine can detect aspirin-induced platelet dysfunction.⁵¹ Clopidogrel effect could not be measured by the ADP channel in testing of cardiology patients.⁵² Both channels, ADP and epinephrine, accurately detect platelet dysfunction in von Willebrand's disease and in uraemia. In cardiac surgical patients, Slaughter and colleagues⁶³ demonstrated that the PFA-100 closure time has only a high negative predictive value and thus might help in identifying patients who are unlikely to need platelet transfusions to reduce bleeding. Its positive predictive value is low and thus it is not very useful in transfusion algorithms to direct transfusion therapy as many 'false-positive' patients would be transfused. If 21 63 75 Positive predictive value has been difficult to attain with many POC tests of platelet function. Cammerer and colleagues have prospectively studied a group of cardiac surgical patients using observational measurement of platelet function to predict bleeding. They reported that thromboelastography (discussed later) was a better predictor of bleeding than the PFA-100, however if the PFA-100 ADP test was used in addition to thromboelastography, the predictive accuracy was enhanced.

VerifyNow[®]

The VerifyNow system (Accumetrics, San Diego, CA, USA) is a POC turbidimetric-based optical detection system that measures agonist-induced agglutination of whole blood. A mixing chamber contains the platelet agonist [thrombin receptor-activating peptide (TRAP), arachidonic acid, or ADP, and fibrinogen-coated beads]. After anticoagulated whole blood is added to the mixing chamber, platelets are activated if they are responsive to the agonist. The activated GPIIb/IIIa receptors on the platelets bind to adjacent platelets via the fibringen on the beads and cause agglutination of the blood and the beads. Light transmittance through the chamber is measured and increases as agglutination increases, much like standard aggregometry. Anti-thrombotic drug effects reduce agglutination (measured by light transmittance) and thus the degree of platelet inhibition can be quantified. Direct pharmacological block of GPIIb/IIIa receptors with a GPIIb/IIIa antagonist is detected with a very high accuracy using this device and the TRAP agonist. More recent cartridges using arachidonic acid as the agonist have been developed that can accurately assess aspirin-induced platelet dysfunction. Through inhibition of arachidonic acid, indirect prevention of GPIIb/IIIa expression is accomplished. The anti-platelet effects of clopidogrel can also be measured using a VerifyNow cartridge that incorporates ADP as the agonist. 43 52 Each of these drug effects can be measured using the appropriate cartridge of the VerifyNow device.74

Platelet Works®

Platelet Works (Helena Laboratories, Beaumont, TX, USA) is a whole-blood assay that uses the principle of the platelet count ratio to assess platelet reactivity. The instrument is a Coulter counter that compares platelet counts in a standard ethylenediaminetetraacetic acid tube with platelet counts in a citrate tube after aggregation with either ADP or collagen. When blood is added to these agonist tubes, platelets activate, adhere to the tube, and are effectively eliminated from the platelet count. The ratio of the

activated platelet count to the non-activated platelet count is a function of reactivity of the platelets. Early investigation indicates that this assay correlates well with standard platelet aggregometry and is capable of measuring the platelet dysfunction induced by GPIIb/IIIa receptor inhibitors and clopidogrel.^{8 40} Although the disadvantage of Platelet Works is that it is not well studied, investigations show that it is capable of measuring the platelet dysfunction that accompanies CPB. ⁴⁹

The real value in testing the platelet response to a specific agonist is derived from the measure of specific platelet defects that accompany anti-thrombotic drug therapy. Many of these tests utilize small doses of agonists that are sensitive to drug therapy but are not sufficient to challenge platelet function that is more severely compromised. When platelet function is overtly deranged, such as after CPB, a potent agonist is necessary to determine whether the platelet can respond. This potent agonist is usually thrombin and is the 'natural' platelet agonist that is used in the visco-elastic tests of clot formation. Thus, the visco-elastic tests, thromboelastography and thromboelastometry, have been most frequently used in transfusion algorithms for bleeding patients as described below.

Visco-elastic tests of clot formation

Sonoclot

The Sonoclot Analyzer (Sienco Inc., Wheat Ridge, CO, USA) is a test of the visco-elastic properties of blood that provides accurate information on the entire haemostasis process including coagulation factors, fibrin gel formation, clot retraction (platelet function), and fibrinolysis. This device consists of a tubular probe that oscillates up and down within a blood sample. The viscous force of the blood creates impedance to the ultrasonic vibrating probe as it clots, which is converted to an output signal. This electronic signal is processed by a microcomputer and is reported as the Clot Signal. The Sonoclot Analyzer reports these properties by graphically recording the dynamics of clot formation as a Sonoclot Signature and also yields quantitative results. The Sonoclot Signature is the plotted values of the Clot Signal value vs time. The quantitative results include a lag period (SonACT) that corresponds to activated clotting time (ACT) and a wave that occurs as a result of cross-linkage of fibrin (Clot RATE). Other parameters in the tracing indicate platelet-fibrin binding, fibrin formation, and clot retraction. Clot retraction is a measure of platelet activity and its quantitative parameter is the time to peak. Haemostasis abnormalities including platelet dysfunction, factor deficiencies, anticoagulant effects, hyperfibrinolysis, and hypercoagulable states can be detected using the Sonoclot. In addition, Sonoclot analysis has been successfully used for diagnosing and treating platelet dysfunction and bleeding disorders after CPB. 20 46 51

Thromboelastograph

The thromboelastograph (TEG), invented in 1948, is another test of the visco-elastic properties of blood that examines the time of initiation through acceleration, control, and eventual lysis. Initially used for coagulation monitoring during liver transplantation, the TEG has found applications in cardiovascular surgery, obstetric anaesthesia, and trauma anaesthesia. 5 22 34 35 77 A small amount of blood (0.36 ml) is placed in an oscillating cuvette and a piston is lowered into the blood sample. The cuvette oscillates at an arc angle of 4° 45 min. As the blood begins to clot, the elastic force exerted on the piston is translated to a signature tracing (thromboelastogram; Fig. 1) that reveals information about fibrin formation, platelet-fibrin interactions, platelet clot strength, and fibrinolysis. With the current disposables, an activator is needed because the onset to coagulation varies, and the time to clot formation can conveniently be accelerated so that the test is useful in POC settings. Celite, kaolin, or tissue factor have all been used to activate the TEG.30 72

There are five parameters to the TEG tracing that measure different stages of clot development: R, K, α angle, maximum amplitude (MA), and MA60 (Fig. 2). In addition, clot lysis indices are measured at 30 and 60 min after MA (LY30 and LY60). Normal values vary depending on the type of activator used. The R value is a

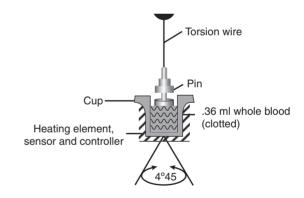


Fig 1 The thromboelastograph uses whole blood in an oscillating cuvette into which a piston is lowered. The movement of the cuvette is translated to an oscillograph tracing as the blood thickens and exerts a force on the piston. This generates the signature thromboelastographic tracing.

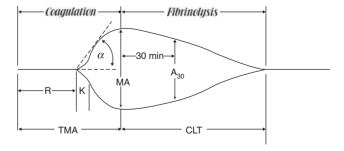


Fig 2 The signature thromboelastograph tracing is shown here with parameters labelled. See text for further details.

measure of clotting time (CT) which is the period of time from the start of the test to the initial fibrin formation. The K value is the clot kinetics measurement of the speed to reach a specific level of clot strength: the time from beginning of clot formation (the end of R time) until the amplitude reaches 20 mm. The α angle is the angle between the horizontal line in the middle of the TEG tracing and the line tangential to the developing 'body' of the TEG tracing at 2 mm amplitude. The α angle represents the acceleration (kinetics) of fibrin build up and cross-linking (clot strengthening). The MA reflects the ultimate strength of the clot which depends on the number and function of platelets and their interaction with fibrin. The MA is the parameter most frequently measured because it correlates with platelet dysfunction in cardiac surgery. The MA is used as a marker for platelet function and has thus been incorporated into transfusion algorithms used to reduce platelet and other transfusions given to patients after CPB. LY30, or the lysis index at 30 min after MA, is increased with fibrinolysis.

A limitation of the TEG is its inability to detect impairment in platelet function induced by anti-platelet agents. The development of the Platelet Mapping Assay[®] has overcome this shortfall.

TEG can be used to predict bleeding in cardiac surgery. 12 14 37 71 In large retrospective and prospective studies, incorporation of the TEG into clinical decisionmaking has resulted in decreased blood loss and transfusions.3 58 61 68 Spiess and colleagues analysed 1079 patients before and after the introduction of TEG as part of an overall transfusion management strategy. They found significantly less use of all blood and blood components except cryoprecipitate. There was also a significant decrease in the re-exploration rate. However, this study may have been biased by the Hawthorne effect (an improvement in results that may be found just by monitoring a process).66 In a prospective randomized controlled study, Shore-Lesserson and colleagues⁶¹ compared transfusion requirements with 'TEG-based' and 'traditional' protocols in the management of postoperative bleeding. Patients in both groups received the antifibrinolytic epsilon aminocaproic acid (EACA). Although the study showed no significant difference in mediastinal tube drainage between the groups, blood and blood component therapy were significantly less in the 'TEG' than in the 'traditional group'. Royston and von Kier⁵⁸ studied 60 patients who had undergone complex surgery comparing their actual blood and blood product use to predicted usage derived from a TEG-based algorithm. They utilized the TEG R value to determine the quantity of fresh frozen plasma needed to reverse the coagulation defect in the post-cardiac surgical patient (Table 1). Though TEG is the best-studied POC device for use in cardiac surgery, further studies are required to recommend TEG as the standard of care for postoperative transfusion management. 13 32

Table 1 TEG-based transfusion algorithm (from Royston and von Kier⁵⁸)

TEG variable	Implication	Therapy
R>14 and <21 mm R>21 and <28 mm R>28 mm MA <48 mm MA <40 mm Lys30 >7.5%	, 2	One fresh frozen plasma Two fresh frozen plasma Four fresh frozen plasma One pooled platelets Two pooled platelets Aprotinin

TEG modifications

Platelet Mapping Assay®

The development of the Platelet Mapping Assay overcomes some of the shortcomings of the TEG, in that it allows for the thromboelastographic measurement of platelet function in patients on anti-platelet medication. Platelet mapping uses three cuvettes. One incorporates thrombin to activate platelets and overrides the inhibition of other activation pathways such as arachidonic acid, ADP, and GPIIb/IIIa. A second cuvette contains reptilase plus factor XIII to create a fibrinogen clot or a 'thrombin-less' clot. This clot strength will be smaller and will not have the contribution of thrombin-activated platelets. The third cuvette incorporates the fibrinogen clot and adds back ADP or arachidonic acid to stimulate the platelets. The ability of the MA to increase in response to ADP (clopidogrel) or arachidonic acid (aspirin) is a measure of drug-induced platelet inhibition via that particular pathway. This POC test correlates well with the gold standard optical aggregometry.8

Rotational thromboelastometry

Rotational thromboelastometry (ROTEM) (Pentapharm, Munich, Germany) provides a visco-elastic measurement of clot strength in whole blood. A small amount of whole blood (0.3 ml) and coagulation activators are added to a disposable cuvette that is placed in a heated holder. A disposable pin (sensor) fixed on the tip of a rotating shaft is lowered into the blood sample. The loss of elasticity upon clotting affects rotation of the shaft that is detected by the reflection of light on a small mirror attached to the shaft. A detector records the axis rotation over time and this rotation is translated into a graph or thromboelastogram.

The main descriptive parameters derived by ROTEM are: CT, corresponding to the time (s) from the beginning of the reaction to a 2 mm increase in amplitude. This represents the initiation of clotting, thrombin formation, and the start of clot polymerization. Clot formation time, the time (s) between an increase in amplitude from 2 to 20 mm. This identifies the fibrin polymerization and stabilization of the clot with platelets and factor XIII. Maximum clot firmness (MCF), the MA (mm) reached in the tracing, which correlates with platelet count, platelet function, and the

concentration of fibrinogen. Alpha (α) angle, the tangent to the clotting curve through the 2 mm point. Maximum lysis, the ratio of the lowest amplitude after MCF to the MCF. Maximum velocity (maxVel), the maximum of the first derivative of the clot curve. Time to maximum velocity (t-maxVel), the time from the start of the reaction until maxVel is reached. The area under curve, defined as the area under the velocity curve, that is, the area under the curve ending at a time point that corresponds to MCF.

ROTEM is approved for use in coagulation monitoring in Europe and its use and familiarity are highest there. In a recent study by Spalding and colleagues⁶⁴ comparing transfusion rates before and after implementation of a ROTEM-based transfusion algorithm, ROTEM[®]-guided coagulation management was useful in the choice of the appropriate therapeutic option in the bleeding patient. This reduced costs by avoiding administration of fresh frozen plasma, cryoprecipitate, and platelet concentrates.⁶⁴ Its use in cardiac surgery and in transfusion algorithms is likely to be similar to that of TEG. ROTEM is currently under consideration as a coagulation monitoring device by the US Food and Drug Administration.

Impact Cone and Plate(let) Analyzer

In the Impact Cone and Plate(let) Analyzer (CPA; DiaMed Cressier, Switzerland), whole blood is exposed to uniform shear by the spinning of a cone in a standardized cup. This allows for platelet function testing under conditions that mimic physiological blood flow, thus achieving the most accurate and authentic pattern of platelet function. After automated staining, platelet adhesion to the cup is evaluated by image analysis software. The success of the CPA in screening for congenital primary haemostasis abnormalities and in testing platelet response to GPIIb/IIIa antagonists, aspirin, and clopidogrel has been demonstrated. Recent studies suggest that the CPA is a useful tool for testing perioperative platelet function and might help predict postoperative blood loss. Experience with this instrument is limited as it has just become commercially available.

POC testing of heparin effect

Heparin and anticoagulation for CPB

CPB perturbs multiple aspects of haemostatic function. Contact between blood and the artificial circuit surface can activate coagulation. Optimal anticoagulation during CPB is necessary to prevent platelet activation and antagonize coagulation to prevent microvascular clots. The most common anticoagulant in clinical use is heparin because it is easy to dose, administer, measure, and reverse. Heparin works by activating antithrombin III (ATIII). Therefore, appropriate levels of ATIII are necessary for heparin to be effective. Activated ATIII inactivates thrombin and other

proteases involved in blood clotting, most notably factor Xa.³¹ For decades, most cardiac surgery programmes utilized empiric heparin dosing starting with a bolus based on a patient's weight (300 U kg⁻¹) and subsequent interval dosing. Empiric dosing has since been replaced by the monitored use of heparin.

Heparin monitoring

Activated clotting time

The ACT is the most commonly used functional POC test to measure heparin anticoagulation. The ACT is an automated variation of the Lee-White CT that uses an activator such as celite or kaolin to activate clotting in a test tube. Early tests used whole blood placed in a warmed test tube with diatomaceous earth as an activator. The tubes were tilted back and forth manually until evidence of clot appeared. Currently, the two most commonly used ACT devices are the Hemochron (International Technidyne Inc., Edison, NJ, USA) and the Hemotec (Medtronic Hemotec, Parker, CO, USA). The Hemochron system consists of a precision aligned magnet within a test tube and a magnet detector located within the well. Whole blood is added to a test tube containing an activator (celite, kaolin, glass beads, or a combination of these) and placed in the well. As clot begins to form, the magnet is lifted within the tube displacing the magnet from the magnet detector. The CT is the time the clot takes to displace the magnet at a given distance. The Hemotec device uses a two-chamber cartridge containing kaolin as an activator. Blood (0.4 ml) is placed into each chamber and a daisy-shaped plunger increases and decreases in the chamber. The formation of clot slows the rate of descent of the plunger, and the decrease in velocity of the plunger is detected by a photooptical system that signals the end of the test. Each ACT analyser is consistent in its ability to reproducibly measure the CT using its specific methodology. There are intrinsic biases built into some of the measurement devices, but repeatability within a given device is high.²

ACT monitoring of heparinization has been criticized because of its high variability. 42 47 50 54 The main limiting factor is that it correlates poorly with anti-Xa measures of heparin activity, or with heparin concentration during CPB as a result of hypothermia and haemodilution. This is especially true of paediatric patients whose consumption of heparin is increased. 45 Other factors altering ACT include thrombocytopenia, the presence of platelet inhibitors, platelet membrane receptor antagonists, and the use of the antifibrinolytic aprotinin (celite only). Blood loss and transfusion requirements in patients undergoing CPB can be reduced with more accurate control of heparin anticoagulation and its reversal.

Cascade POC System

A completely different technology for measuring the effect of heparin is used by the Cascade POC System (Helena). This system uses disposable cards with celite

activator to measure heparin activity. This variant of the ACT is called the heparin management test (HMT). This card contains paramagnetic iron oxide particles that move in response to an oscillating magnetic field. When clot formation occurs, movement of the iron oxide particles is decreased. This system is capable of measuring prothrombin time (PT) and activated partial thromboplastin time (aPTT), which will be discussed below. The suitability of this platform for monitoring of ACT during cardiac surgery has been demonstrated in a variety of clinical studies. 53 57 HMT correlates well with anti-Xa heparin activity in CPB and is less variable than standard ACT. In a comparison with ACT, the coefficients of variation were similar between the tests at baseline but were three times higher for the ACT during heparinization. This degree of agreement with plasma anti-Xa measurements has not been demonstrated universally in patients undergoing CPB.¹⁹

Individual heparin dosing

In vitro techniques have been introduced to measure patient dose-response to heparin. These assays measure the sensitivity to a known quantity of heparin and generate a doseresponse curve that enables calculation of the heparin dose required to attain the target anticoagulation. Blood loss and transfusion requirements in cardiac surgical patients can be reduced with more accurate control of heparin anticoagulation and its reversal.^{27 32} Similarly, a protamine doseresponse curve can be generated using an in vitro sample with a known quantity of protamine, thus enabling protamine dosing to be based only upon the level of circulating heparin. The Hemochron RxDX (International Technidvne Corp.) system is an ACT-based heparin dose-response (HDR) assay. Heparin requirement is measured by the heparin response test, and the required protamine dose is measured by the protamine response test. 33 A separate test, the protamine dose assay measures residual-free heparin in the blood. Using this system, other investigators have been able to significantly lower protamine doses, and some have reported significantly reduced transfusions and chest tube drainage in the group that received individualized dosing with RxDx. Another in vitro heparinized dose-response assay is the Hepcon (Medtronic) HDR, which constructs a three-point HDR curve using the baseline, 1.5, and 2.5 IU ml⁻¹ heparin. From this curve, extrapolation to the desired ACT or heparin concentration yields the indicated dose of heparin. These dose-response assays are used less frequently than weight-based heparin dosing since the latter technique is faster, less expensive, and extremely safe when monitored. It is not clear that individualized heparin dosing alone, in the absence of individualized protamine dosing, affects perioperative blood loss and transfusions in cardiac surgery.

POC monitoring of coagulation status

Current transfusion guidelines for blood-component therapy strongly recommend results of PT and aPTT to

guide administration of fresh frozen plasma and cryoprecipitate. Because of the lag time in obtaining results from a central laboratory, many decisions regarding transfusion of blood products are based on clinical judgement.²³ Several POC coagulation analysers are currently available. The former Thrombolytic Assessment System (Pharmaetics Inc., Raleigh, NC, USA), now the Cascade POC (Helena), which was previously discussed for its ability to measure heparin by the HMT, also measures PT and aPTT. The sample is added to a cartridge containing paramagnetic iron oxide particles that oscillate in a magnetic field as described. Specific activating reagents are used for each assay, including rabbit brain thromboplastin for the PT, aluminium magnesium silicate for aPTT, and celite for HMT. The blood moves by capillary action and mixes with paramagnetic iron oxide particles and reagent within the testing chamber. The decreased movement of the particles is detected optically as the sample clots and the result is displayed as time (s) and as international normalized ratio (INR) for PT. The CoaguChekProDM monitor (Roche Diagnostics, Manheim, Germany) uses a wholeblood sample added to a test cartridge that contains a soybean activator and phospholipids. As the sample clots, a laser optically monitors the decrease in blood flow, and the resultant CT is displayed in seconds for PT and aPTT, and as a ratio for INR. This device has been studied in cardiac surgical patients, and the PT result compared favourably with laboratory plasma-based assays at most perioperative time points. The aPTT result also correlated with plasma-based samples but was slightly less accurate and had a significant bias. 48 These same authors used a previous version of this monitor to define normal vs abnormal aPTT and PT values after CPB to predict which patients were more likely to have bleeding. A very important application of this POC device has been its use in transfusion algorithms to direct transfusion of fresh frozen plasma and platelets after cardiac surgery. 18 Despotis and colleagues⁹ randomly assigned patients with microvascular bleeding to a transfusion algorithm using only the platelet count and the POC PT/aPTT device. The control group received transfusion therapy using standard laboratory testing. In their algorithm, bleeding patients with abnormal coagulation tests were transfused fresh frozen plasma, and bleeding patients with near normal coagulation tests were given platelet pharmacological enhancers or platelet concentrates. Overall, the patients who were treated using the POC transfusion algorithm received significantly fewer allogeneic transfusions than the control group. 9 10

Conclusions

The benefits of POC testing in surgical patients include rapid turnaround times and specific measurements of haemostasis defects that can direct therapy. The use of specific tests that can be used at the bedside has enabled

Table 2 POC transfusion algorithm outcome studies

Author	Surgery type	Study type	Patients	Outcome
Despotis and colleagues ¹¹	Cardiac	Prospective	362	Algorithm decreased transfusion and bleeding
Avidan and colleagues ³	Cardiac	Prospective	102	Two algorithms decreased transfusion
Spiess and colleagues ⁶⁶	Cardiac	Retrospective	1079	TEG use decreased transfusion
Shore- Lesserson and colleagues ⁶¹	Cardiac	Prospective	102	TEG algorithm decreased transfusion
Nuttall and colleagues ⁴⁸	Cardiac	Prospective	836	Algorithm decreased transfusion and bleeding
Capraro and colleagues ⁶	Cardiac	Prospective	1412	Algorithm increased platelet transfusion, no difference in bleeding
Royston and von Kier ⁵⁸	Cardiac and heart transplant	Prospective	60	TEG algorithm decreased transfusion

the incorporation of these tests into transfusion algorithms that more directly address the haemostatic problem and allow treatment using fewer allogeneic blood products (Table 2). The tests reviewed here are those that have been studied in the laboratory, in patients, or both and have proven accurate in monitoring haemostatic function. Many have been used in algorithms to treat bleeding patients. There is a major need for anti-platelet drug testing in surgical patients. The ability to detect preoperative platelet inhibition due to clopidogrel or other anti-thrombotic agents will allow stratification and rational management of surgical patients. The use of platelet function monitoring in transfusion algorithms for bleeding patients will increase as we continue to perform more complex surgeries in cardiovascular patients.

References

- I Ak K, Isbir CS, Tetik S, et al. Thromboelastography-based transfusion algorithm reduces blood product use after elective CABG: a prospective randomized study. J Card Surg 2009; 24: 404–10
- 2 Akl BF, Vargas GM, Neal J, Robillard J, Kelly P. Clinical experience with the activated clotting time for the control of heparin and protomine therapy during cardiopulmonary bypass. J Thorac Cardiovasc Surg 1980; 79: 97–102
- 3 Avidan MS, Alcock EL, Da Fonseca J, et al. Comparison of structured used of routine laboratory tests or near-patient assessment with clinical judgement in the management of bleeding after cardiac surgery. Br J Anaesth 2004; 92: 178–86
- 4 Bjork I, Lindahl U. Mechanism of the anticoagulant action of heparin. Mol Cell Biochem 1982; 48: 161–82
- 5 Cammerer U, Dietrich W, Rampf T, Braun SL, Richter JA. The predictive value of modified computerized thromboelastography

- and platelet function analysis for postoperative blood loss in routine cardiac surgery. *Anesth Analg* 2003; **96**: 51–7
- 6 Capraro L, Kuitunen A, Salmenperä M, Kekomäki R. On-site coagulation monitoring does not affect hemostatic outcome after cardiac surgery. Acta Anaesthesiol Scand 2001; 45: 200-6
- 7 Craft RM, Chavez JJ, Breese SJ, Wortham DC, Cohen E, Carroll RC. A novel modification of the Thromboelastograph assay, isolating platelet function, correlates with optical platelet aggregation. J Lab Clin Med 2004; 143: 301–9
- 8 Craft RM, Chavez JJ, Snider CC, Muenchen RA, Carroll RC. Comparison of modified Thromboelastograph and Plateletworks whole blood assays to optical platelet aggregation for monitoring reversal of clopidogrel inhibition in elective surgery patients. J Lab Clin Med 2005: 145: 309–15
- 9 Despotis GJ, Joist JG, Goodnough LT. Monitoring of haemostasis in cardiac surgical patients: impact of point-of-care testing on blood loss and transfusion outcomes. Clin Chem 1997; 42: 1684–96
- 10 Despotis GJ, Renna M, Eby C. Risks associated with bleeding and transfusion: rationale for the optimal management of bleeding after cardiac surgery. Eur J Anaesthesiol 2007; 24: 15–36
- 11 Despotis GJ, Santoro SA, Spitznagel E, et al. Prospective evaluation and clinical utility of on-site monitoring of coagulation in patients undergoing cardiac operation. J Thorac Cardiovasc Surg 1994: 107: 271–9
- 12 Dorman BH, Spinale FG, Bailey MK, Kratz JM, Roy RC. Identification of patients at risk for excessive blood loss during coronary artery bypass surgery: thromboelastography vs. coagulation screen. Anesth Analg 1993; 76: 694–700
- 13 Dunning J, Versteegh M, FAbbri A, et al. Guideline on antiplatelet and anticoagulation management in cardiac surgery. Eur J Cardiothorac Surg 2008; 34: 73–92
- 14 Essell JH, Martin TJ, Salinas J, Thompson JM, Smith VC. Comparison of thromboelastography to bleeding time and standard coagulation tests in patients after cardiopulmonary bypass. J Cardiothorac Vasc Anesth 1993; 7: 410–5
- 15 Faraday N, Guallar E, Sera VA, et al. Utility of whole blood hemostatometry using the clot signature analyzer for assessment of haemostasis in cardiac surgery. Anesthesiology 2002; 96: 1115-22
- 16 Fattorutto M, Pradier O, Schmatrz D, Ickx B, Barvais L. Does platelet function analyzer (PFA-100) predict blood loss after cardiopulmonary bypass? Br J Anaesth 2003; 90: 692–3
- 17 Ferraris A, Ferraris SP, Saha SP, et al. Perioperative blood transfusion and blood conservation in cardiac surgery: the Society of Thoracic Surgeons and the Society of Cardiovascular Anesthesiologists Clinical Practice Guideline. Ann Thorac Surg 2007; 83: S27-86
- 18 Ferring M, Reber G, de Moerloose P, et al. Point of care and central laboratory determinations of the aPTT are not interchangeable in surgical intensive care patients. Can J Anaesth 2001; 48: 1155-60
- 19 Flom-Halvorsen HI, Ovrum E, Abdelnoor M, et al. Assessment of heparin anticoagulation: comparison of two commercially available methods. Ann Thorac Surg 1999; 67: 1012–6, discussion 6–7
- 20 Forestier F, Belisle S, Contant CH, Harel F, Janvier G, Hardy JF. Reproducibility and interchangeability of the Thromboelastograph, Sonoclot, and Hemochron activated coagulation time in cardiac surgery. Can J Anaesth 2001; 48: 902–10
- 21 Forestier F, Coiffic A, Mouton C, Ekouevi D, Chene G, Janvier G. Platelet function point-of-care tests in post-bypass cardiac surgery: are they relevant? Br J Anaesth 2002; 89: 715–21

- 22 Ganter MT, Hofer CK. Coagulation monitoring: current techniques clinical use of viscoelastic point-of-care coagulation devices. Anesth Analg 2008; 106: 1366-75, abstract
- 23 Gelb AB, Roth RI, Levin J, et al. Changes in blood coagulation during and following cardiopulmonary bypass. Am J Clin Pathol 1996: 106: 87–99
- 24 Gerrah R, Brill A, Tshori S, Lubetsky A, Merin G, Varon D. Using cone and plate(let) analyzer to predict bleeding in cardiac surgery. Asian Cardiovasc Thorac Ann 2006; 14: 310–5
- 25 Goodnough LT, Johnston MF, Toy PT. The variability of transfusion practice in coronary artery bypass surgery. Transfusion Medicine Academic Award Group. J Am Med Assoc 1991; 265: 86–90
- 26 Gorlinger K, Jambor C, Hanke AA, et al. Perioperative coagulation management and control of platelet transfusion by point-of-care platelet function analysis. Transfus Med Hemother 2007; 34: 396–411
- 27 Gravlee GP, Rogers AT, Dudas LM, et al. Heparin management protocol for cardiopulmonary bypass influences post-operative heparin rebound but not bleeding. Anesthesiology 1992; 76: 393-401
- 28 Gurbel PA, Bliden KP, DiChiara J, et al. Evaluation of dose-related effects of aspirin on platelet function: results from the Aspirin-Induced Platelet Effect (ASPECT) study. Circulation 2007; 115: 3156-64
- 29 Hall TS, Sines JC, Spotnitz AJ. Hemorrhage related reexploration following open heart surgery: the impact of pre-operative and postoperative coagulation testing. *Cardiovasc Surg* 2002; 10: 146–53
- **30** Hartert H, Schaeder JA. The physical and biological constraints of thromboelastography. *Biorheology* 1962; 1: 31–9
- 31 Hirsh J. Heparin. N Engl | Med 1991; 324: 1565-74
- **32** Jobes DR, Aitken GL, Shaffer GW. Increased accuracy and precision of heparin and protamine dosing reduces blood loss and transfusion in patients undergoing primary cardiac operations. *J Thorac Cardiovasc Surg* 1995; **110**: 36–45
- 33 Jobes DR, Schwartz AJ, Ellison N, Andrews R, Ruffini RA, Ruffini JJ. Monitoring heparin anticoagulation and its neutralization. *Ann Thorac Surg* 1981; 31: 161–6
- 34 Kang YG, Martin D, Marquez J, et al. Intraoperative changes in blood coagulation and thrombelastographic monitoring in liver transplantation. Anesth Analg 1985; 64: 888–97
- 35 Kaufman CR, Dwyer KN, Cruz JD, Dols SJ, Trask AL. Usefulness of thromboelastography in assessment of trauma patient coagulation. J Trauma 1997; 42: 716–20
- 36 Kestin AS, Valeri CR, Khuri SF, et al. The platelet function defect of cardiopulmonary bypass. Blood 1993; 82: 107–17
- 37 Koster A, Kukucka M, Fischer T, Hetzer R, Kuppe H. Evaluation of post-cardiopulmonary bypass coagulation disorders by differential diagnosis with a multichannel modified thromboelastogram: a pilot investigation. J Extra Corpor Technol 2001; 33: 153–8
- 38 Kundu SK, Heilmann EJ, Garcia C, Davidson RM, Ostgaard RA. Description of an in vitro platelet function analyzer: PFA-100. Semin Thromb Hemost 1995; 21: 106-12
- 39 Lasne D, Fiemeyer A, Chatellier G, Chammas C, Baron JF, Aiach M. A study of platelet function with a new analyzer using high shear stress (PFA 100) in patients undergoing coronary artery bypass graft. Thromb Haemost 2000; 84: 794–9
- 40 Lau WC, Waskell LA, Watkins PB, et al. Atorvastatin reduces the ability of clopidogrel to inhibit platelet aggregation: a new drugdrug interaction. Circulation 2003; 107: 32-7
- 41 Lawrence L. Detailed diagnoses and procedures for patients discharged from short stay hospitals: United States, 1984. Hyattsville, MD: National Center for Health Statistics, 1986: 169. [Vital and

- Health Statistics. Series 13: Data from the National Health Survey; no. 86 (DHHS publication no. PHS 86-1747).]
- **42** Lefemine AA, Lewis M. Activated clotting time for control of anticoagulation during surgery. *Am Surg* 1985; **51**: 274–8
- 43 Lordkipanidze M, Pharand C, Nguyen TA, et al. Assessment of VerifyNow P2Y12 assay accuracy in evaluating clopidogrelinduced platelet inhibition. Ther Drug Monit 2008; 30: 372–8
- 44 Maquelin KN, Berckmans RJ, Nieuwland R, et al. Disappearance of glycoprotein lb from the platelet surface in pericardial blood during cardiopulmonary bypass. J Thorac Cardiovasc Surg 1998; 115: 1160-5
- **45** Miller BE, Mochizuki T, Levy JH, et al. Predicting and treating coagulopathies after cardiopulmonary bypass in children. *Anesth Analg* 1997; **85**: 1196–202
- 46 Miyashita R, Kuro M. Evaluation of platelet function by Sonoclot analysis compared with other hemostatic variables in cardiac surgery. Anesth Anal 1998; 87: 1228–33
- 47 Niinikoski J, Laato M, Laaksonen V, Jalonen J, Inberg MV. Use of activated clotting time to monitor anticoagulation during cardiac surgery. Scand | Thorac Cardiovasc Surg 1984; 18: 57–61
- **48** Nuttall GA, Oliver WC, Santrach PJ, et al. Efficacy of a simple intraoperative transfusion algorithm for nonerythrocyte component utilization after cardiopulmonary bypass. *Anesthesiology* 2001; **94**: 773–81, discussion 5–6A.
- **49** Ostrowsky J, Foes J, Warchol M, Tsarovsky G, Blay J. Plateletworks platelet function test compared to the thromboelastograph for prediction of postoperative outcomes. *J Extra Corpor Technol* **2004**; **36**: 149–52
- **50** Ottesen S, Stormorken H, Hatteland K. The value of activated clotting time in monitoring heparin therapy during extracorpeal circulation. *Scand J Thorac Cardiovasc Surg* 1984; 18: 123–8
- **51** Paniccia R, Antonucci E, Gori AM, et al. Comparison of different methods to evaluate the effect of aspirin on platelet function in high-risk patients with ischemic heart disease receiving dual antiplatelet treatment. Am | Clin Pathol 2007; **128**: 143–9
- **52** Paniccia R, Antonucci E, Gori AM, et al. Different methodologies for evaluating the effect of clopidogrel on platelet function in high-risk coronary artery disease patients. J Thromb Haemost 2007; **5**: 1835–8
- 53 Papaconstantinou C, Radegran K. Use of the activated coagulation time in cardiac surgery. Effects on heparin-protamine dosages and bleeding. Scand J Thorac Cardiovasc Surg 1981; 15: 213–5
- 54 Preiss DU, Schmidt-Bleibtreu H, Berguson P, Metz G. Blood transfusion requirements in coronary artery surgery with and without activated clotting time (ACT) technique. Klin Wochenschr 1985; 63: 252–6
- 55 Rinder CS, Mathew JP, Rinder HM, Bonan J, Ault KA, Smith BR. Modulation of platelet surface adhesion receptors during cardiopulmonary bypass. *Anesthesiology* 1991; 75: 563–708
- 56 Rochon AG, Shore-Lesserson L. Coagulation monitoring. Anesthesiol Clin North Am 2006; 24: 839–56
- 57 Roth JA, Cukingnan RA, Scott CR. Use of activated coagulation time to monitor heparin during cardiac surgery. Ann Thorac Surg 1979; 28: 69–72
- 58 Royston D, von Kier S. Reduced haemostatic factor transfusion using heparinase-modified thrombelastography during cardiopulmonary bypass. Br J Anaesth 2001; 86: 575–8
- 59 Samama CM, Ozier Y. Near-patient testing of haemostasis in the operating theatre: An approach to appropriate use of blood in surgery. Vox Sanguinis 2003; 84: 251–5
- **60** Savion N, Varon D. Impact-the cone and plate(let) analyzer: testing platelet function and anti-platelet drug response. *Pathophysiol Haemost Thromb* 2006; **35**: 83–8

- 61 Shore-Lesserson L, Manspeizer HE, DePerio M, Francis S, Vela-Cantos F, Ergin MA. Thromboelastography-guided transfusion algorithm reduces transfusion in complex cardiac surgery. Anesth Analg 1999; 88: 312–9
- 62 Skubas NJ, Despotis GJ. Optimal management of bleeding complications after cardiac surgery. Semin Cardiothorac Vasc Anesth 2001; 5: 217–28
- 63 Slaughter TF, Sreeram G, Sharma AD, et al. Reversible shear-mediated platelet dysfunction during cardiac surgery as assessed by the PFA-100 platelet function analyzer. Blood Coagul Fibrinolysis 2001: 12: 85-93
- 64 Spalding GJ, Hartrumpf M, Sierig T, Oesberg N, Kirschke CG, Albes JM. Cost reduction of perioperative coagulation management in cardiac surgery: value of 'bedside' thrombelastography (ROTEM). Eur | Cardiothorac Surg 2007; 31: 1052-7
- 65 Spiess BD. Transfusion of blood products affects outcome in cardiac surgery. Semin Cardiothorac Vasc Anesth 2004; 8: 267–81
- 66 Spiess BD, Gillies BS, Chandler W, Verrier E. Changes in transfusion therapy and reexploration rate after institution of a blood management program in cardiac surgical patients. J Cardiothorac Vasc Anesth 1995; 9: 168–73
- 67 Spiess BD, Royston D, Levy JH, et al. Platelet transfusions during coronary artery bypass graft surgery are associated with serious adverse outcomes. *Transfusion* 2004: 44: 1143–8
- 68 Spiess BD, Tuman KJ, McCarthy RJ, DeLaria GA, Schillo R, Ivankovich AD. Thromboelastography as an indicator of post-cardiopulmonary bypass coagulopathies. J Clin Monit 1987; 3: 25–30

- **69** Steiner ME, Despotis GJ. Transfusion algorithms and how they apply to blood conservation: The high-risk cardiac surgical patient. Hematol Oncol Clin North Am 2007; **21**: 177–84
- 70 Tanaka K, Takao M, Yada I, Yuasa H, Kusagawa M, Deguchi K. Alterations in coagulation and fibrinolysis associated with cardio-pulmonary bypass during open heart surgery. J Cardiothorac Anesth 1989; 3: 181–8
- 71 Ti LK, Cheong KF, Chen FG. Prediction of excessive bleeding after coronary artery bypass graft surgery: the influence of timing and heparinase on thromboelastography. J Cardiothorac Vasc Anesth 2002; 16: 545–50
- 72 Tuman KJ, McCarthy RJ, Djuric M, Rizzo V, Ivankovich AD. Evaluation of coagulation during cardiopulmonary bypass with a heparinase modified thromboelastographic assay. *J Cardiothorac Vasc Anesth* 1994; 8: 144–9
- 73 Tuman KJ, Spiess BD, McCarthy RJ, Ivankovich AD. Comparison of viscoelastic measures of coagulation after cardiopulmonary bypass. Anesth Analg 1989; 69: 69-75
- 74 van Werkum JW, Harmsze AM, Elsenberg EH, et al. The use of the VerifyNow system to monitor antiplatelet therapy: a review of the current evidence. Platelets 2008; 19: 479–88
- 75 Whaba A, Sander S, Birnbaum DE. Are in-vitro platelet function tests useful in predicting blood loss following open heart surgery? *Thorac Cardiovasc Surg* 1998; 46: 228–31
- 76 Woodman RC, Harker LA. Bleeding associated with cardiopulmonary bypass. Blood 1990; 76: 1680–97
- 77 Zuckerman L, Cohen E, Vagher JP, Woodward E, Caprinia JA. Comparison of thromboelastography with common coagulation tests. *Thromb Haemost* 1981; 46: 752–6